

Antiproliferative Potential of Extracts from *Kappaphycus* Seaweeds on HeLa Cancer Cell Lines

(Potensi Antiproliferatif Ekstrak Rumpai Laut *Kappaphycus* ke atas Titisian Sel Kanser HeLa)

TIEK YING LAU, DELBORA FENNY VITTAL, CASSANDRA SZE YII CHEW
& WILSON THAU LYM YONG*

ABSTRACT

A review of the current literature indicates that natural seaweeds are an excellent source of bioactive compounds with antioxidant, antimicrobial and antitumor properties. In the present study, 90% methanolic, 70% acetonetic and aqueous extracts from *Kappaphycus alvarezii* (strains Crocodile, Giant and Brown) and *Kappaphycus striatum* were used to inhibit the growth of HeLa cell lines. MTS assay was carried out to determine the proliferation of HeLa cells in the presence of different seaweed extracts. Both 500 $\mu\text{g/mL}$ of aqueous and methanolic extracts from *K. striatum* demonstrated highest anti-proliferative activity against HeLa cells with cell growth inhibition of 53.5 and 43.7%, respectively. Treatment with the aqueous extracts from three strains of *K. alvarezii* did not show any growth inhibition against HeLa cell lines. The acetonetic extract of *Kappaphycus* seaweeds exhibited a very poor cell growth inhibition with inhibitory activity observed under the treatment of 300 to 500 $\mu\text{g/mL}$ of *K. alvarezii* strain Brown only. Further studies are suggested to identify and purify the specific anti-tumoral compounds for potential use in cancer therapy.

Keywords: Antiproliferation; growth inhibition; *Kappaphycus alvarezii*; *Kappaphycus striatum*

ABSTRAK

Suatu kajian kepustakaan semasa menunjukkan bahawa rumpai laut semula jadi adalah sumber komponen bioaktif yang sangat baik dengan aktiviti antioksidan, antimikrobial dan antitumor. Dalam kajian ini, ekstrak metanol 90%, aseton 70% dan akueus daripada *Kappaphycus alvarezii* (strain Buaya, Giant dan Brown) dan *Kappaphycus striatum* telah digunakan untuk merencat pertumbuhan titisian sel HeLa. Asai MTS telah dijalankan untuk mengkaji pertumbuhan sel HeLa dalam kehadiran pelbagai ekstrak rumpai laut. Kedua-dua 500 $\mu\text{g/mL}$ ekstrak akueus dan metanol daripada *K. striatum* menunjukkan aktiviti antiproliferasi yang tertinggi terhadap sel HeLa masing-masing dengan perencatan pertumbuhan sel sebanyak 53.5 dan 43.7%. Rawatan dengan ekstrak akueus daripada tiga strain *K. alvarezii* tidak menunjukkan perencatan pertumbuhan terhadap titisian sel HeLa. Ekstrak aseton daripada rumpai laut *Kappaphycus* mempamerkan perencatan pertumbuhan sel yang lemah dengan aktiviti perencatan hanya boleh diperhatikan dengan rawatan 300 hingga 500 $\mu\text{g/mL}$ daripada *K. alvarezii* strain Brown sahaja. Kajian lanjutan adalah dicadangkan untuk mengenal pasti dan menuliskan komponen khusus antitumor untuk pembangunan terapi kanser.

Kata kunci: Antiproliferasi; *Kappaphycus alvarezii*; *Kappaphycus striatum*; perencatan tumbuhan

INTRODUCTION

Seaweeds are considered to be a source of bioactive compounds as they are able to produce a variety of secondary metabolites characterized by a broad spectrum of biological activities. Compounds with cytostatic, antiviral, anthelmintic, antifungal and antibacterial activities have been detected in green, brown and red algae (Lindequist & Schweder 2001; Newman et al. 2003). More recently, seaweeds are reported to be a rich source of antioxidant compounds (Duan et al. 2006; Kuda et al. 2005; Lim et al. 2002). For example, chlorophylls, carotenoids, tocopherol derivatives such as vitamin E and related isoprenoids, which are structurally related to plant-derived antioxidants, were found in some marine organisms including seaweeds (Takamatsu et al. 2003). Antioxidants in biological systems have multiple functions, including defense against oxidative

damage and participating in the major signaling pathways of cells. Besides, some compounds from the seaweeds have antibacterial activities with potential use as mosquito control agents. Extracts from *Eucheuma denticulatum* have exhibited antibacterial activity on Gram positive organisms including *Staphylococcus aureus* and *Streptococcus pyogenes* (Al-Haj et al. 2009).

Seaweeds also contain bioactive substances with great potential as antitumoral drugs, which lead to emerging interests in the biomedical research in seaweeds (Michio et al. 1984; de Sousa et al. 2007). Several species of seaweeds are rich sources of polysaccharides and glycoproteins with immune-stimulant, anticancer or antiviral activity (Abdel-Fattah et al. 1974; de Sousa et al. 2007; Michio et al. 1984; Nishino et al. 1989; Smit 2004). Certain algae have long been used in traditional Chinese herbal medicine in cancer

treatment (Yamamoto et al. 1984). Red and green algae have been shown to demonstrate protective effects against mammary, intestinal and skin carcinogenesis (Yuan & Walsh 2006). Zandi et al. (2010) reported that cold water extract of red alga, *Gracilaria corticata*, possessed biological activity against tumor cells replication. In recent years, much attention has been focused on fucoidan, a sulphated polysaccharide derived from brown seaweeds. Recent studies evidenced that fucoidan has strong antitumor activity and exhibited important roles against human cancer cell lines (Ly et al. 2005; Matsuda et al. 2010). Fucoidan was found to be able to suppress the growth of tumor cells *in vivo* and activate the immune system against tumors (Itoh et al. 1993; Maruyama et al. 2003; Noda et al. 1990; Usui et al. 1980; Yamamoto et al. 1984; Zhuang et al. 1995).

The two red seaweed species, *K. alvarezii* and *K. striatum*, which are extensively distributed in Sabah, have been uncovered as a novel source for a variety of compounds such as dietary fibers, vitamin C, α -tocopherol, minerals, fatty acid and protein (Matanjun et al. 2008). However, there is limited information about their biological activity on cancer cell growth inhibition. The objectives of this study were to screen and evaluate the anti-proliferative activities of crude methanolic, acetonetic and aqueous extracts of selected strains of *K. alvarezii* and *K. striatum*. The information compiled during the course of this study can be of use for further development of cancer therapy.

MATERIALS AND METHODS

SAMPLES PREPARATION

Kappaphycus alvarezii (strains Crocodile, Giant and Brown) and *Kappaphycus striatum* were collected from Semporna, Sabah. The samples were washed with fresh water and dried at room temperature for 1 week. The dried seaweed samples were separately milled and subjected to compound extractions. For aqueous extraction, dry powder of seaweed was macerated with de-ionized water and filtered through cotton wool and Whatman (No. 1) filter paper to remove debris. The filtrate was lyophilized using freeze dryer for 3 days. For each of extraction using 90% methanol and 70% acetone, approximately 100 g of powdered seaweed samples were extracted using a soxhlet apparatus. The methanol and acetone were purchased from Sigma-Aldrich (St. Louis, MO, USA). About 500 mL of each solvent was used to carry out the extraction in soxhlet apparatus for a period of 24–72 h until the solvent becomes colorless at $65 \pm 2^\circ\text{C}$. The solvent was evaporated using a rotary vacuum evaporator to make the final volume one-fourth of the original volume. The methanolic, acetonetic and aqueous extracts were stored in -10°C for further analysis of anti-proliferative assay in triplicate.

CELL LINE AND CULTURE CONDITION

HeLa Cancer Cell Line CCL-23™ was purchased from American Type Culture Collection (ATCC®, USA). Cells

were seeded and grown in RPMI (Roswell Park Memorial Institute) media. They were maintained in 12.5 cm^3 BD Falcon™ cell culture flask (California, USA) at 37°C in a humidified atmosphere with 5% CO_2 . The RPMI medium was replaced once every two days and passaging was performed to maintain the adherent cell lines.

GROWTH INHIBITION ASSAY

In order to observe the seaweed extracts responsiveness, a cell proliferation assay was carried out. The inhibition effects of methanolic, acetonetic and aqueous extracts on the growth of HeLa cells were evaluated *in vitro* by the MTS assay. This method relies on the ability of dehydrogenase enzymes in the metabolically active cells to convert 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium or MTS to a formazan precipitate. The Cell Titer 96® Aqueous Non-Radioactive Cell Proliferation Assay purchased from Promega (Madison, USA) was used to determine the cell proliferation of HeLa cells in the presence of different types of extracts (methanolic, acetonetic and aqueous) at different concentrations (50, 100, 200, 300, 400 and 500 $\mu\text{g}/\text{mL}$). The different concentrations of each extracts were prepared from the stock solutions by serial dilution.

A known number of HeLa cells (10^4) were incubated in 96-well plates in a volume of 200 μL of culture medium and permitted to adhere for 24 h before addition of test compounds. About 100 μL of different concentrations (50, 100, 200, 300, 400 and 500 $\mu\text{g}/\text{mL}$) of each extracts (methanolic, acetonetic and aqueous) were added to the cells. After 48 h of exposure, the cells were washed with 100 μL of phosphate-buffered saline (PBS) and replaced with fresh medium. Approximately 20 μL of CellTiter 96® AQ_{ueous} One Solution Reagent was added into each well of the 96-well assay containing the samples in 100 μL of culture medium. The plates were incubated at 37°C in a humidified atmosphere with 5% CO_2 . Following incubation for 4 h, the plates were read with SPECTRAMax M2 ROM (Molecular Devices) microplate reader at absorbance of 490 nm. The experiments were performed twice in triplicate. The results were evaluated by comparing the absorbance of the treated cells with the absorbance of wells containing cell treated by the solvent control. Conventionally, cell viability was estimated to be 100% in the solvent control.

DATA ANALYSES

Percentage of cell growth inhibition versus extracts concentration was calculated according to Patel et al. (2009) as follow:

$$\text{Percentage of cell growth inhibition} = 100 - [(A-B)/(C-B)] \times 100,$$

where A is the absorbance of sample; B is the absorbance of blank and C is the absorbance of control.

RESULTS

MTS assay was carried out to investigate the inhibition effects of methanolic, acetic and aqueous extracts of *Kappaphycus* seaweeds on the growth of HeLa cells and the results are represented in Figures 1 to 4. Six different concentrations (50, 100, 200, 300, 400 and 500 $\mu\text{g}/\text{mL}$) of each extract (methanolic, acetic and aqueous) were applied. Figure 1 shows the percentage of growth inhibition against 90% methanolic, 70% acetic and aqueous extracts of *K. alvarezii* strain Crocodile. Among the three types of extracts, only methanolic extract from 200 to 500 $\mu\text{g}/\text{mL}$ showed obvious anti-proliferative activity against the HeLa cells. The highest percentage (31.7%) of growth inhibition was observed with the treatment using 300 $\mu\text{g}/\text{mL}$ of methanolic extract. This was followed by the treatment using methanolic extracts at 400, 500 and 200 $\mu\text{g}/\text{mL}$ with 24.6, 16.5 and 4.0% of growth inhibition, respectively. Treatments with 50 and 100 $\mu\text{g}/\text{mL}$ methanolic extracts and all acetic and aqueous extracts did not show any growth inhibition but they promoted growth of the treated cells instead.

For the treatment with different extracts from *K. alvarezii* strain Giant, only methanolic extract at 400 $\mu\text{g}/\text{mL}$ demonstrated growth inhibition (8.5%) as summarized in Figure 2. Methanolic extract with other concentrations and all the acetic and aqueous extracts from *K. alvarezii* strain Giant did not show any cell growth inhibition. While for the treatment with *K. alvarezii* strain Brown extracts, the highest growth inhibition (30.4%) was observed with 50 $\mu\text{g}/\text{mL}$ of methanolic extract, as shown in Figure 3. Increment of methanolic extract concentrations resulted in decrement of growth inhibition as seen with 100 and 200 $\mu\text{g}/\text{mL}$ of extracts demonstrated 7.9 and 5.9% of growth inhibition, respectively. On the other hand, only acetic extract from this strain demonstrated positive anti-proliferative activity against the HeLa cells as compared with the other two strains (Crocodile and Giant). Increment

of acetic extract from 300 to 500 $\mu\text{g}/\text{mL}$ had parallel increment in the inhibition percentage from 17.5 to 29.7%.

Figure 4 shows the percentage of cell growth inhibition against concentrations of methanolic, acetic and aqueous extracts of *K. striatum*. The results indicated that all the methanolic and aqueous extracts have positive inhibition on the HeLa cell lines. Treatment with the aqueous extract ranging from 50 to 500 $\mu\text{g}/\text{mL}$ exhibited concentration dependent anti-proliferative activity against HeLa cells with 17.8 to 53.5% of cell growth inhibition. Whereas, the inhibition effect of methanolic extract on cells growth ranged from 6.2 to 43.7%. All acetic extracts from *K. striatum* did not inhibit, but promote, cell growth.

DISCUSSION

Marine algae contain many unidentified useful components and physiologically active substances. Studies on bioactivity of marine algae against cancer cell lines have been reported in previous researches, where the findings have brought great promise to the development of cancer treatment activities (Albano et al. 1990; Berlinck et al. 1996). Some studies involved general extractions of seaweeds while others applied extraction of specific metabolites such as carotene, bromophenols and carrageenan (Ly et al. 2005; Xu et al. 2004). In the present study, 90% methanolic extracts, 70% acetic extracts and aqueous extracts of *K. alvarezii* and *K. striatum* were studied for their potential to inhibit the growth of HeLa cell lines. The most effective concentration to inhibit cell growth was found to be 500 $\mu\text{g}/\text{mL}$ of aqueous extract of *K. striatum* followed by 500 $\mu\text{g}/\text{mL}$ of methanolic extract of same species, with 53.5 and 43.7% of growth inhibition, respectively. These differences in antitumor activities may be attributed to their different molecular weights, charge characteristics and monosaccharide distributions (Dias et al. 2005).

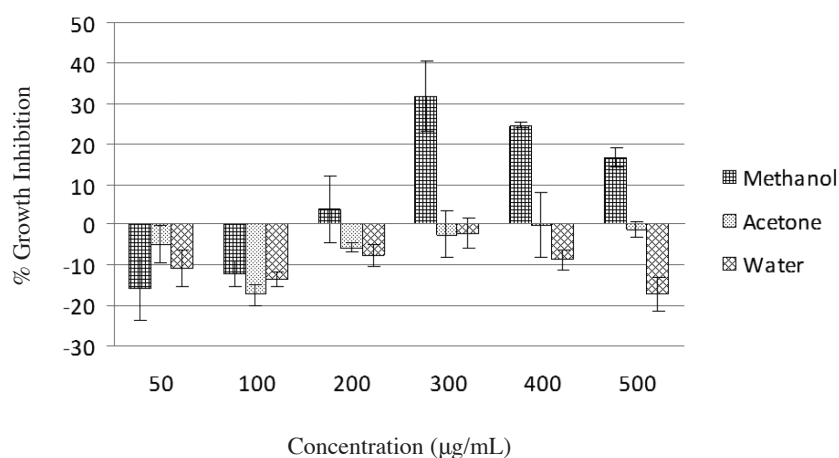


FIGURE 1. Percentage of growth inhibition of HeLa cell lines in the presence of 90% methanolic, 70% acetic and aqueous extracts of *K. alvarezii* strain Crocodile. Data points show the mean \pm SE for a minimum of three experiments

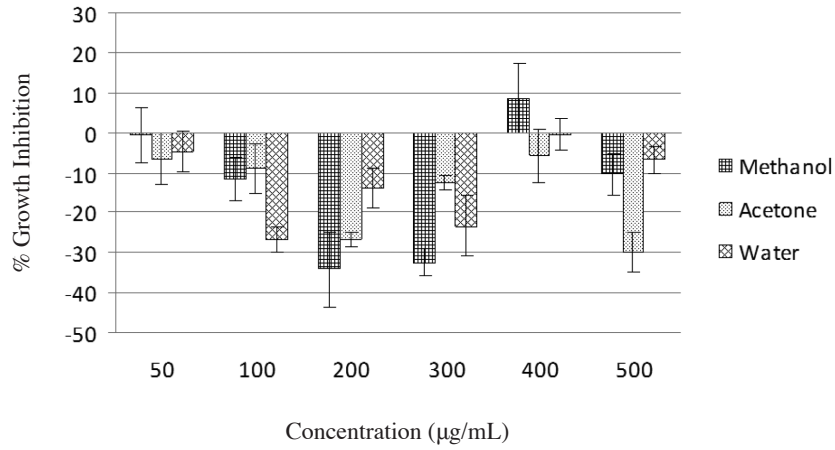


FIGURE 2. Percentage of growth inhibition of HeLa cell lines in the presence of 90% methanolic, 70% acenotic and aqueous extracts of *K. alvarezii* strain Giant. Data points show the mean ± SE for a minimum of three experiments

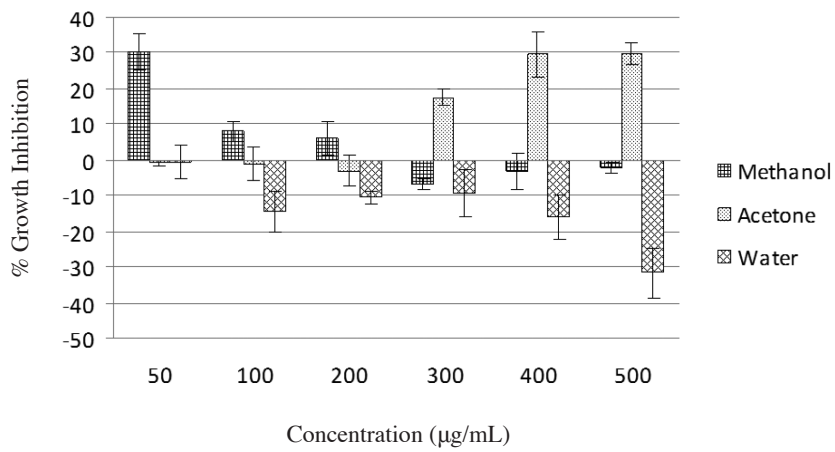


FIGURE 3. Percentage of growth inhibition of HeLa cell lines in the presence of 90% methanolic, 70% acenotic, and aqueous extracts of *K. alvarezii* strain Brown. Data points show the mean ± SE for a minimum of three experiments

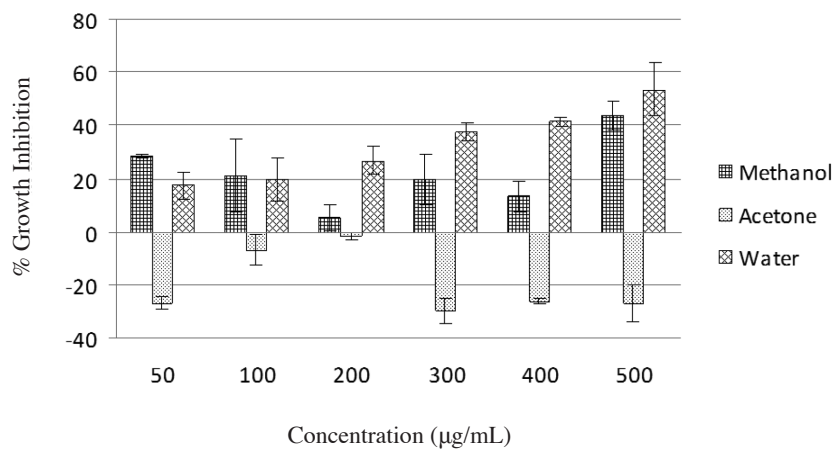


FIGURE 4. Percentage of growth inhibition of HeLa cell lines in the presence of 90% methanolic, 70% acenotic and aqueous extracts of *K. striatum*. Data points show the mean ± SE for a minimum of three experiments

Previous studies reported that alcoholic extracts from plant samples exhibited several bioactivities such as adaptogenic, anti-inflammatory, anticonvulsant, sedative, androgenic and immunopromoting activities (Xu et al. 1992). This might be the reason why methanolic extracts from *Kappaphycus* seaweeds generally showed positive growth inhibition to the HeLa cell lines as compared with acetonic and aqueous extracts. Studies by Shao et al. (1996) also reported that alcoholic extract from asparagus shoots exhibited antitumor activities and Singh et al. (1992) reported their fruit to be the source of bile-stimulating agent. Reports from World Intellectual Property Organization (2010) also indicated that methanolic extracts from various seaweed species have demonstrated cytotoxic effect on human cancer cell lines including HeLa, MCF-7 and MDA-MB-231. Alcohol is found to be effective to extract active compounds such as biophenols, lipids, saccharides, minerals and small peptides due to their polarity. The potential bioactive compounds in seaweed may interact with special cancer associated receptors or cancer specific molecules to trigger the mechanisms leading to cancer cell death.

Previous researches show that acetone-water mixtures are good solvent systems for the extraction of polar antioxidants (Lu & Yeap Foo 1999; Luximon-Ramma et al. 2005; Sun 2002). Literature also describes that acetone and water extracts of plant flowers presented the best total phenolic content (Liu et al. 2009). Nyenje and Ndip (2011) suggested that an organic solvent, in particular, acetone is a good solvent as it extracts more active compounds from plant material. Flavonoids and steroids have also been reported to be extracted using acetone according to Abdulmalik et al. (2011) and Eloff (1998). Besides, van Slambrouck et al. (2007) demonstrated that crude aqueous extracts of *L. tridentata* (Creosote Bush) and *J. communis* L. (Juniper Berry) have significantly decreased the growth of MCF-7/AZ breast cancer cells. Traditional medicines are often prepared by water extraction, but water-soluble impurities present challenges for conventional isolation methods, such as chromatography or crystallization (Bart 2011). Water preferentially extracts polar compounds but they need some special post treatment such as ion exchange or caustic wash for further purification (Jones & Kingkorn 2006).

Further studies are suggested to identify the specific anti-tumoral compounds in the targeted extracts. Purification can be carried out to obtain the bioactive compounds for the development of cancer therapy. Besides, identification of specific metabolites such as carotene, bromophenols and carrageenan from seaweeds is also recommended for the discovery of potential anti-proliferative or anticancer compounds.

ACKNOWLEDGEMENTS

The authors wish to thank the Ministry of Education (MOE), Malaysia for funding the research under the Fundamental Research Grant Scheme (FRG0201-SG-1/2010).

REFERENCES

- Abdel-Fattah, A.F., Hussein, M.M. & Salem, H.M. 1974. Studies of the purification and some properties of sargasan, a sulphated heteropolysaccharide from *Sargassum linifolium*. *Carbohydrate Research* 33: 9-17.
- Abdulmalik, I.A., Sule, M.I., Musa, A.M., Yaro, A.H., Abdullahi, M.I., Abdulkadir, M.F. & Yusuf, H. 2011. Isolation of steroids from acetone extract of *Ficus iteophylla*. *British Journal of Pharmacology and Toxicology* 2: 270-272.
- Albano, R.M., Pavao, M.S.G., Mourao, P.A.S. & Mulloy, B. 1990. Structural studies of a sulfated L-galactan from *Styela plicata* (Tunicate): Analysis of the Smith-degraded polysaccharide. *Carbohydrate Research* 208: 163-174.
- Al-Haj, N.A., Mashan, N.I., Shamsudin, M.N., Mohamad, H., Vairappan, C.S. & Sekawi, Z. 2009. Antibacterial activity in marine algae *Eucheuma denticulatum* against *Staphylococcus aureus* and *Streptococcus pyogenes*. *Research Journal of Biological Sciences* 4: 519-524.
- Bart, H.J. 2011. Extraction of natural products from plants - An introduction. In *Industrial Scale Natural Products Extraction*. 1st ed., edited by Bart, H.J. & Pilz, S. Weinheim: Wiley-VCH Verlag GmbH & Co. KGaA. pp. 1-25.
- Berlinck, R.G.S., Ogawa, C.A., Almeida, A.M.P., Sanchez, M.A.A., Malpezzi, E.L.A., Costa, L.V., Hajdu, E. & de Freitas, J.C. 1996. Chemical and pharmacological characterization of halitoxin from *Amphimedon viridis* (Porifera) from the southeastern Brazilian coast. *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology* 115: 155-163.
- de Sousa, A.P.A., Torres, M.R., Pessoa, C., de Moraes, M.O., Filho, F.D.R., Alves, A.P.N.N. & Costa-Lotufo, L.V. 2007. *In vivo* growth-inhibition of Sarcoma 180 tumor by alginates from brown seaweed *Sargassum vulgare*. *Carbohydrate Polymers* 69: 7-13.
- Dias, P.F., Siqueira, J.M., Vendruscolo, L.F., de Jesus Neiva, T., Gagliardi, A.R., Maraschin, M. & Ribeiro-do-Valle, R.M. 2005. Antiangiogenic and antitumoral properties of a polysaccharide isolated from the seaweed *Sargassum stenophyllum*. *Cancer Chemotherapy and Pharmacology* 56: 436-446.
- Duan, X.J., Zhang, W.W., Li, X.M. & Wang, B.G. 2006. Evaluation of antioxidant property of extract and fractions obtained from a red alga, *Polysiphonia urceolata*. *Food Chemistry* 95: 37-43.
- Eloff, J.N. 1998. Which extractant should be used for the screening and isolation of antimicrobial components from plants? *Journal of Ethnopharmacology* 60: 1-8.
- Itoh, H., Noda, H., Amano, H., Zhuang, C., Mizuno, T. & Ito, H. 1993. Antitumor activity and immunological properties of marine algal polysaccharides, especially fucoidan, prepared from *Sargassum thunbergii* of phaeophyceae. *Anticancer Research* 13: 2045-2052.
- Jones, W.P. & Kingkorn, A.D. 2006. Extraction of plant secondary metabolites - Natural Products Isolation. In *Methods in Biotechnology*. vol. 20, 2nd ed., edited by Sarker, S.D., Latif, Z. & Gray, A.I. Totowa, New Jersey: Humana Press. pp. 323-351.
- Kuda, T., Tsunekawa, M., Goto, H. & Araki, Y. 2005. Antioxidant properties of four edible algae harvested in the Noto Peninsula, Japan. *Journal of Food Composition and Analysis* 18: 625-633.
- Lim, S.N., Cheung, P.C.K., Ooi, V.E.C. & Ang, P.O. 2002. Evaluation of antioxidative activity of extracts from a brown seaweed, *Sargassum siliquastrum*. *Journal of Agricultural and Food Chemistry* 50: 3862-3866.
- Lindequist, U. & Schweder, T. 2001. Marine biotechnology. In

- Biotechnology*, vol. 10, edited by Rehm, H.J. & Reed, G. Weinheim: Wiley-VCH, pp. 441-484.
- Liu, S.C., Lin, J.T., Wang, C.K., Chen, H.Y. & Yang, D.J. 2009. Antioxidant properties of various solvent extracts from lychee (*Litchi chinensis* Sonn.) flowers. *Food Chemistry* 114: 577-581.
- Lu, Y. & Yeap Foo, L. 1999. The polyphenol constituents of grape pomace. *Food Chemistry* 65: 1-8.
- Luximon-Ramma, A., Bahorun, T., Crozier, A., Zbarsky, V., Datla, K.P., Dexter, D.T. & Aruoma, O.I. 2005. Characterization of the antioxidant functions of flavonoids and proanthocyanidins in Mauritian black teas. *Food Research International* 38: 357-367.
- Ly, B.M., Buu, N.Q., Nhut, N.D., Thinh, P.D., Thi, T. & Van, T. 2005. Studies on fucoidan and its production from Vietnamese brown seaweeds. *Asean Journal for Science and Technology Development (AJSTD)* 22: 371-380.
- Matanjun, P., Mohamed, S., Mustapha, N.M., Muhammad, K. & Ming, C.H. 2008. Antioxidant activities and phenolics content of eight species of seaweeds from north Borneo. *Journal of Applied Phycology* 20: 367-373.
- Maruyama, H., Tamauchi, H., Hashimoto, M. & Nakano, T. 2003. Antitumor activity and immune response of Mekabu fucoidan extracted from Sporophyll of *Undaria pinnatifida*. *In vivo* 17: 245-249.
- Matsuda, Y., Teruya, K., Matsuda, S., Nakano, A., Nishimoto, T., Ueno, M., Niho, A., Yamashita, M., Eto, H., Katakura, Y. & Shirahata, S. 2010. Anti-cancer effects of enzyme-digested fucoidan extract from seaweed Mozuku. *Animal Cell Technology: Basic & Applied Aspects* 16: 295-300.
- Michio, F., Noriko, I., Ichiro, Y. & Terukazu, N. 1984. Purification and chemical and physical characterization of an antitumor polysaccharide from the brown seaweed *Sargassum fulvellum*. *Carbohydrate Research* 125: 97-106.
- Newman, D.J., Cragg, G.M. & Snader, K.M. 2003. Natural products as source of new drugs over the period 1981-2002. *Journal of Natural Products* 66: 1022-1037.
- Nishino, T., Yokoyama, G., Dobashi, K., Fujihara, M. & Nagumo, T. 1989. Isolation, purification and characterization of fucose-containing sulphated polysaccharides from the brown seaweed *Ecklonia kurome* and their blood-anticoagulant activities. *Carbohydrate Research* 186: 119-129.
- Noda, H., Amano, H., Arashima, K. & Nisizawa, K. 1990. Antitumor activity of marine algae. *Hydrobiologia* 204-205: 577-584.
- Nyenje, M. & Ndip, R.N. 2011. *In-vitro* antimicrobial activity of crude acetone extract of the stem bark of *Combretum molle* against selected bacterial pathogens. *Journal of Medicinal Plants Research* 5: 5315-5320.
- Patel, S., Gheewala, N., Suthar, A. & Shah, A. 2009. *In-vitro* cytotoxicity activity of *Solanum nigrum* extract against HeLa cell line and Vero cell line. *International Journal of Pharmacy and Pharmaceutical Sciences* 1: 38-46.
- Shao, Y., Chin, C.K., Ho, C.T., Ma, W., Garrosp, S.A. & Huang, M.T. 1996. Anti-tumor activity of the crude saponins obtained from asparagus. *Cancer Letters* 104: 31-36.
- Singh, A.K., Hussain, A., Srivastava, G.N., Misra, L.N., Gupta, M.M., Virmani, O.P., Popli, S.P. & Abraham, Z. 1992. *Dictionary of Indian Medicinal Plants*. Lucknow, India: Central Institute of Medicinal and Aromatic Plants (CIMAP).
- Smit, A.J. 2004. Medicinal and pharmaceutical uses of seaweed natural products: A review. *Journal of Applied Phycology* 16: 245-262.
- Sun, S.G. 2002. Reply to the comments on the paper by Zheng M.S. and Sun S.G. entitled 'In situ FTIR spectroscopic studies of CO adsorption on electrodes with nanometer-scale thin films of ruthenium in sulphuric acid solutions' by Pecharrómán, C., Cuesta, A. & Gutiérrez, C. *Journal of Electroanalytical Chemistry* 529: 155-158.
- Takamatsu, S., Hodges, T.W., Rajbhandari, I., Gerwick, W.H., Hamann, M.T. & Nagle, G. 2003. Marine natural products as novel antioxidant prototypes. *Journal of Natural Products* 66: 605-608.
- Usui, T., Asari, K. & Mizuno, T. 1980. Isolation of highly purified "Fucoidan" from *Eisenia bicyclis* and its anticoagulant and antitumor activities. *Agricultural and Biological Chemistry* 44: 1965-1966.
- van Slambrouck, S., Daniels, A.L., Hooten, C.J., Brock, S.L., Jenkins, A.R., Ogasawara, M.A., Baker, J.M., Adkins, G., Elias, E.M., Agustin, V.J., Constantine, S.R., Pullin, M.J., Shors, S.T., Kornienko, A. & Steelant, W.F.A. 2007. Effect of crude aqueous medicinal plant extracts on growth and invasion of breast cancer cells. *Oncology Reports* 17: 1487-1492.
- Xu, J.P., Xu, R.S. & Li, X.Y. 1992. Four new cycloartane saponins from *Curculigo orchioides*. *Planta Medica* 58: 208-210.
- Xu, N., Fan, X., Yan, X. & Tseng, C.K. 2004. Screening marine algae from China for their antitumor activities. *Journal of Applied Phycology* 16: 451-456.
- Yamamoto, I., Takahashi, M., Suzuki, T., Seino, H. & Mori, H. 1984. Antitumor effect of seaweeds. IV. Enhancement of antitumor activity by sulfation of a crude fucoidan fraction from *Sargassum kjellmanianum*. *The Japanese Journal of Experimental Medicine* 54: 143-151.
- Yuan, Y.V. & Walsh, N.A. 2006. Antioxidant and antiproliferative activities of extracts from a variety of edible seaweeds. *Food and Chemical Toxicology* 44: 1144-1150.
- Zandi, K., Tajbakhsh, S., Nabipour, I., Rastian, Z., Yousefi, F., Sharafiah, S. & Sartavi, K. 2010. *In vitro* antitumor activity of *Gracilaria corticata* (a red alga) against Jurkat and molt-4 human cancer cell lines. *African Journal of Biotechnology* 9: 6787-6790.
- Zhuang, C., Itoh, H., Mizuno, T. & Ito, H. 1995. Antitumor active fucoidan from the brown seaweed, umitoranoo (*Sargassum thunbergii*). *Bioscience, Biotechnology, and Biochemistry* 59: 563-567.
- Tiek Ying Lau, Cassandra Sze Yii Chew & Wilson Thau Lym Yong*
Biotechnology Research Institute
Universiti Malaysia Sabah, Jalan UMS
88400 Kota Kinabalu, Sabah
Malaysia
- Delbora Fenny Vittal
School of Science and Technology
Universiti Malaysia Sabah, Jalan UMS
88400 Kota Kinabalu, Sabah
Malaysia

*Corresponding author; email: wilsonyg@ums.edu.my

Received: 10 March 2013

Accepted: 24 April 2014